

Original Article

The therapeutic potential of Nelumbo Nucifera against oxidative damage induced by Carbon Tetrachloride in rats.

Sagheera Bibi & Lubna Naz

Department of Physiology, University of Karachi, Karachi-Pakistan.



Doi: 10.29052/IJEHSR.v10.i2.2022.213-219

Corresponding Author Email:

lunaz@uok.edu.pk Received 23/10/2021 Accepted 10/01/2022 First Published 25/05/2022



© The Author(s). 2022 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/)

Abstract

Background: Herbal treatment has paid much attention in the world over the past decades. Nelumbo nucifera is used all over the world as a medicinal plant. It is an aquatic perennial plant belonging to the Nelumbonaceae family. The protective effects of Nelumbo nucifera are due to the active ingredients present in it. The study was conducted to evaluate the protective effects of Nelumbo nucifera seeds against hepatotoxicity caused by carbon tetrachloride (CCI4) in rats.

Methodology: Eighteen Wistar Albino rats were allotted into three groups (n=6), control group, CCl4 treated group given CCl4 0.8 ml/kg body weight subcutaneously twice a week, CCl4 + N. Nucifera treated group given CCl4 0.8 ml/kg body weight 2 times a week subcutaneously and also given Nelumbo nucifera extract (200 mg/kg body weight) orally through gavage daily. After 28 days of the experiment, serum samples were taken for estimation of enzyme activities and liver tissue samples were collected for evaluation of antioxidant enzyme levels.

Results: It was observed that liver weight increased in the CCl4 treated group while liver weight decreased in CCl4 + N. Nucifera treated group. CCl4 administration significantly raises the transaminase levels such as Aspartate transaminase (AST) (p<0.01), Aspartate transaminase (ALT) (p<0.05), and Alkaline phosphatase (ALP) (p<0.05) whereas CCl4 + N. Nucifera treated group serum transaminases are reduced. Antioxidant activities such as Catalase (p>0.05), Glutathione (GSH) (p>0.05) and Superoxide dismutase (SOD) (p=0.05) decreased in the CCl4 treated group. Significant elevation of antioxidant enzymes; Catalase (p<0.05), GSH (p<0.05) and SOD (p>0.05) in CCl4 + N. Nucifera treated group. Serum Malondialdehyde (MDA) concentration (p<0.05) significantly raised in the CCl4 hepatotoxic group compared to the control group. A decreased MDA levels were observed in the CCl4 + N. Nucifera treated group, indicating its potential to counteract the harmful effects of CCl4.

Conclusion: This study suggests that Nelumbo nucifera seeds have overcome CCl4-induced hepatotoxicity by their antioxidant effects.

Keywords

Herbal Treatment, Nelumbo Nucifera, Carbon Tetrachloride, Oxidative Stress.



Introduction

Liver, a vital organ in the body, accounts approximately 2-3% of body weight. Liver is at risk of deterioration due to a variety of factors such as microorganisms, metabolic products, circulatory materials as well as toxins. Pathologic problems like liver triglycerides and fatty liver have a key role in the developmental of various metabolic disorders like hypertension, diabetes mellitus, dyslipidemia, insulin resistance and obesity indicates the contribution of non-alcoholic fatty liver disease (NAFLD) management in the health promotion¹. Because drug metabolism is an important function of the liver, the liver is at higher risk of being damaged by drugs than other organs. The second major fatal cancer in the world is liver cancer². Liver injury induced by the drug leads to hepatitis, liver fibrosis, failure and consequent death³.

Herbs or their extracts have gained considerable worldwide concentration due to their considerable effectiveness in the treatment and prevention of certain diseases^{4,5}. In Asian countries, herbs have been used since thousands of the year as a cure for the disease. A recent study documented that 65% of the European countries relies on the herbal remedies⁶. One thousand one hundred or more medicines have been identified to cause liver toxicity⁷. According to researchers, herbal medicines have significant protective effects due to the presence of compounds such as alkaloids, flavonoids, saccharides and phenylpropanoids⁸.

Nelumbo nucifera (lotus) has been used as herbal medicine, functional food as well as vegetable over 2000 years⁹. It is an aquatic plant cultivated mostly in Asia and Africa. Flowers, leaves, rhizomes, seeds, almost all parts of the plant have beneficial protective effects. Seeds and rhizomes are the most widely used parts¹⁰. Studies have been investigated for the health benefits of using lotus seeds due to the presence of alkaloids. Lotus leaves are used for treatment of obesity in China¹¹. Studies revealed that Nelumbo nucifera seeds have antiamnesic¹², anti-tumor¹³, anti-oxidant¹⁴ and anti-inflammatory^{15,16} and hepatoprotective¹⁷ effects. It is believed that the protective effects of Nelumbo nucifera are attributed to its active ingredients such

as alkaloids and flavonoids¹⁸. Nelumbo nucifera seeds contain protein 25%, carbohydrate 65%, ash 4%, crude fibre 3-4%, moisture 8-10% and 388 cal/100 g of energy. It contains minerals such as calcium, potassium, sodium, copper, phosphorus, magnesium, manganese, selenium, zinc and iron¹⁹.

To develop new therapeutic agents require more effort as pharmacotherapeutic options for liver diseases are limited²⁰. Therefore the aim of the present study is to investigate the protective effects of Nelumbo Nucifera seeds against oxidative damage induced by carbon tetrachloride in rats.

Methodology

Plant Material

Juna market in Karachi was selected to collect Nelumbo nucifera seeds. The dried seeds were ground into powder using a grinder. Then 200 grams of powder was soaked in 1000 ml D.H2O. It was well mixed and tightly covered with polythene paper. The resulting extract was then placed in shaking incubator at room temperature for 24 hrs. Whatman No 1 filter paper was used for extract filtration and filtrate so obtained was stored in a closed container at 2°C²¹.

Experimental Animals

Male and female Albino wistar rats (n=18) weighing between 120-200 gms were obtained from the International Centre for Chemical and Biological Sciences, Karachi. The animals were housed in airconditioned animal house at department of physiology, university of Karachi. They were placed in clean plastic cages with acclimatization with laboratory environment 1 week prior to start the experiment. Standard rodent diet and ad libitum water was given to rats. Body weights of the animals were recorded at specific time interval throughout the experiment.

Experimental Design

Rats were assigned into three different groups (n=6). Group I served as control group. Group II animals were administered CCI4 (0.8 ml/kg b.w) 2 times a week subcutaneously. Group III was given Nelumbo nucifera extract (200 mg/kg b.w) orally

daily along with CCl4 (0.8 ml/kg b.w) subcutaneously 2 times a week for 28 days.

Sample Collection

On the 29th day, animals were decapitated and blood samples were collected in heparin-coated gel tubes. Samples were then subjected to centrifugation for separation of serum & plasma at 3000 rpm for five minutes. Serum samples were then stored for further analysis at 20°C. Liver were excised out from the animals, washed with saline and weighed. Homogenization of liver samples was done to prepare liver homogenate.

Biochemical Assay

Liver function was estimated by measuring serum transaminases. Serum ALT and AST levels were determined according to Reitman and Frankel²². Tietz and associates method was used to analyze

ALP serum activity²³. Antioxidant enzyme activities such as catalase²⁴, glutathione reductase²⁵ and superoxide dismutase²⁶ were evaluated. The level of lipid peroxidation was estimated by the serum MDA concentration as per Okawa et al method²⁷.

Statistical analysis

Data was expressed as mean standard deviation and analyzed using SPSS version 16. One way analysis of variance (ANOVA) was used to compare means of control with experimental groups where p < 0.05 was considered significant.

Results

In present study, it was observed that liver weight was higher in CCl4 treated and CCl4 + N. nucifera treated animals than control group (Table 1).

Table 1: Comparison of liver weights among experimental groups.

	Control	CCI ₄ ¹	CCl ₄ + N. nucifera ^{1,2}
Variable	(n=6)	(n=6)	(n=6)
	Mean ± SD		
Liver Weight (g)	4.75±0.911	7.31±1.00 ^v	7.483±0.626 ^{γ,n,β}

¹Compared with control, ²Compared with CCl4 group. $^{\alpha}p < 0.05$, $^{\beta}p < 0.01$, $^{\gamma}p < 0.001$, $^{\delta}p = 0.05$, $^{n}p > 0.05$.

Transaminases such as AST, ALT and ALP concentrations were assessed and compared (Table 2). Compared to Control, significant increases in AST (p<0.01), ALT (p<0.05) and ALP (p<0.05) levels in CCl4 treated group. In contrast, AST, ALT and ALP levels were decreased in CCl4 + N. nucifera group compared to CCl4 treated, but this change was observed non-significant (p>0.05).

Table 2: Liver enzymes; AST, ALT and ALP levels in experimental groups.

Variable	Control	CCI ₄ ¹	CCl ₄ + N. nucifera ^{1,2}
	(n=6)	(n=6)	(n=6)
		Mean ± SD	
AST (U/L)	10.536± 6.51	28.62±8.09 ^β	16.612±14.355 ^{n,n,n}
ALT (U/L)	3.34±2.11	7.77±4.54°	4.966±2.978 ^{n,n,n}
ALP (U/L)	11.91±11.16	31.98±10.67°	28.115±11.88 ^{α,n,n}

AST-Aspartate Transaminase; ALT-Aspartate Transaminase; ALP-Alkaline Phosphatase.

The activity of antioxidant enzymes was also analyzed in the present study (Table 3). Compared to control, the concentration of Catalase (P>0.05), GSH (P>0.05) and SOD (p=0.05) decreased significantly in CCl4 treated rats. It has been observed that N. nucifera administration significantly increased the levels of catalase (p<0.05)

¹Compared with control, ²Compared with CCl4 group.

 $^{^{\}alpha}p$ <0.05, $^{\beta}p$ <0.01, $^{\gamma}p$ <0.001, $^{\delta}p$ =0.05, ^{n}p >0.05.

and GSH (p<0.05) while SOD concentration increased (p>0.05) non-significantly compared to CCl4 treated group. CCl4 + N. nucifera group showed an increase in the SOD concentration as compared to CCl4 treated.

Table 3: Comparison of Catalase, GSH and SOD levels among experimental groups.

Variable	Control (n=6)	CCI ₄ ¹ (n=6)	CCI4 + N. nucifera ^{1,2} (n=6)	
Catalase (µmol/g tissue)	135.63±2.90	132.55±6.46 ⁿ	$140.93 \pm 3.73^{\alpha,\alpha,n}$	
GSH (unit/g tissue)	89.12±18.64	71.58±22.88 ⁿ	196.83±78.54 ^{α,α,η}	
SOD (unit/g tissue)	20.23±6.14	13.33±3.91 ^δ	16.71±4.63 ^{n,n,n}	

GSH-Glutathione; SOD-Superoxide dismutase

The hepatic MDA level in the experimental groups was evaluated and compared (Table 4). There were significantly elevated MDA levels (p < 0.05) in CCl4 treated group when compared to control. Decreased MDA concentration in CCl4 + N. nucifera group as compared with CCl4 treated group was observed.

Table 4: Comparison of MDA levels among experimental groups.

Variable	Control (n=6)	CCI4 ¹ (n=6)	CCl4 + N. nucifera ^{1,2} (n=6)	
	Mean ± SD			
MDA (μmol/g tissue)	2.310±0.788	6.191±2.08 α	4.480±0.846 β,n,n	

MDA- Malondialdehyde.

¹Compared with control, ²Compared with CCl4 group.

 $^{\alpha}p$ <0.05, $^{\beta}p$ <0.01, $^{\nu}p$ <0.001, $^{\delta}p$ =0.05, ^{n}p >0.05.

Discussion

Xenobiotics are known as hepatotoxic agents that damage intracellular structures such as mitochondria and plasma membrane²⁸. CCl4, a halogenated alkane, is one of the xenobiotics that contribute to hepatic damage through lipid per oxidation. It causes extensive damage to liver tissue such as fibrosis, fatty degeneration and impaired liver function. CCI4 induced liver injury involves transformation of carbon tetrachloride into trichloromethyl radicals that leads to reactive oxygen species (ROS) generation resulting in oxidative damage²⁹. In this study, CCl4 treatment group showed decrease in body weights whereas and an increase in liver weight indicating the liver damage.

Elevation of serum liver enzymes is the prominent marker of liver damage. Accordingly, significant elevations of AST (p<0.01), ALT (p<0.05) and ALP (p<0.05) levels of CCl4 hepatotoxic group were

found in our results. These results indicate cellular leakage, damage of membrane functional integrity and hepatocyte dysfunction as reported previously³⁰. Animals received CCl4 + N. nucifera seeds showed decrease non-significantly in the serum concentrations of AST, ALT and ALP indicates the ability of N. nucifera to counteract the CCl4 effects. The protective effects of N. nucifera are attributed to the various alkaloids present in it.

Oxidative stress affects mitochondrial function by acting directly impairing oxidative phosphorylation. Reactive nitrogen species or reactive oxygen species can provoke mitochondrial membrane permeability transition and mitochondrial DNA deletions resulting in activation of caspases that leads to cell death. Studies reported that the liver diseases linked with oxidative stress are fatty liver, fibrosis, cirrhosis chronic hepatitis and carcinoma³¹. Our result showed a decline in the antioxidant enzymes such

¹Compared with control, ²Compared with CCl4 group.

 $^{^{\}alpha}p$ <0.05, $^{\beta}p$ <0.01, $^{\nu}p$ <0.001, $^{\delta}p$ =0.05, ^{n}p >0.05.

as CAT (p>0.05), SOD (p=0.05) and GSH (p>0.05) in the CCl4 treatment group, which is consistent with previous studies³². Nelumbo nucifera administration enhanced the catalase (p<0.05), GSH (p<0.05) and SOD (p>0.05) concentration in our study that shows the antioxidant effects of Nelumbo nucifera against CCl4 induced liver injury. Similarly, the findings of Sohn et al, resemble with our observation in which liver and kidney peroxidize activity improved by administration of Nelumbo nuciera alcoholic extract³³.

Carbon tetrachloride transformed into CCI3 radical when binds with liver cytochrome P450 that result in initiation of lipid peroxidation³⁴. The secondary metabolites of CCI4 react with proteins and lipids leading alteration of membrane permeability, mitochondria and plasma membrane results in cell damage. During lipid peroxidation secondary metabolites such as 4-hydroxynonenal, hexanal and MDA formed. In our results increased in the MDA levels in CCl4 administered experimental group shows the progressive lipid peroxidation caused by CCI4. Decreased MDA concentration in CCI4 + Nelumbo nucifera treated group indicates the potential of N.nucifera to overcome deleterious effects of CCI4. Studies documented that phytochemicals such as saponins, alkaloids, carbohydrates and polyphenolics found in the Nelumbo nucifera seeds attributed antioxidant potential¹⁰.

Specific doses of herbs used and their active ingredients need further research as a possible natural treatment to limit the progression of liver disease.

Conclusion

The findings of our study suggest that carbon tetrachloride produces hepatotoxicity as it elevates liver serum enzymes and lowers antioxidant concentration that results in oxidative stress. Nelumbo nucifera seeds have hepatoprotective potential by reducing liver enzymes and improve antioxidant levels thus counteracting the harmful effects of CCl4. Further studies are required to explore the protective effects of Nelumbo nucifera

and their active components to limit the liver disease progression.

Conflicts of Interest

The authors have declared that no competing interests exist.

Acknowledgement

We are grateful to the teachers, lab attendants and co-workers of the Department of Physiology, University of Karachi.

Funding

The author(s) received no specific funding for this work

References

- 1. Moore JB. Non-alcoholic fatty liver disease: the hepatic consequence of obesity and the metabolic syndrome. Proc Nutr Soc. 2010;69(2):211-220.
- Chen F, Zhong Z, Tan HY, Guo W, Zhang C, Tan CW, Li S, Wang N, Feng Y. Uncovering the anticancer mechanisms of Chinese herbal medicine formulas: Therapeutic alternatives for liver cancer. Front. Pharmacol. 2020;11:293.
- 3. Hebels DG, Jetten MJ, Aerts HJ, Herwig R, Theunissen DH, Gaj S, Van Delft JH, Kleinjans JC. Evaluation of database-derived pathway development for enabling biomarker discovery for hepatotoxicity. Biomark Med. 2014;8(2):185-200.
- Ghayas H, Sohail S, Iqbal Khan N, Yasmeen G, Naz L, Shamim M. Cardioprotective role of salvia rosmarinus (rosemary) leaves against oxidative stress and in balancing lipid profile in mice. Int. j. endorsing health sci. res. 2021;9(1):61-69.
- Jeong SM, Seo BK, Park YC, Baek YH. A review of complementary and alternative medicine therapies on muscular atrophy: a literature review of in vivo/in vitro studies. eCAM. 2018; Article ID 8654719.
- 6. Singh S, Bajpai M, Mishra P. Herbal Folklore Medication for Liver Disorders. Curr Tradit Med. 2021;7(3):415-433.
- 7. Hoofnagle JH. LiverTox: a website on drug-induced liver injury. Hepatology. 2013;57(3):873-874
- 8. Sajid A, Naz L, Mughal A, Yasmeen G, Khan NI, Sohail S. Antioxidant effects of Moringa oleifera seed oil against oxidative stress induced by alloxan in rats. Int. j. endorsing health sci. res. 2020;8(4):257-264.

- Chen G, Zhu M, Guo M. Research advances in traditional and modern use of Nelumbo nucifera: Phytochemicals, health promoting activities and beyond. Crit Rev Food Sci Nutr. 2019;59(sup1):S189-209.
- Rai S, Wahile A, Mukherjee K, Saha BP, Mukherjee PK. Antioxidant activity of Nelumbo nucifera (sacred lotus) seeds. J Ethnopharmacol. 2006;104(3):322-327.
- 11. Ono Y, Hattori E, Fukaya Y, Imai S, Ohizumi Y. Antiobesity effect of Nelumbo nucifera leaves extract in mice and rats. J Ethnopharmacol. 2006;106(2):238-244.
- 12. Jung HA, Karki S, Kim JH, Choi JS. BACE1 and cholinesterase inhibitory activities of Nelumbo nucifera embryos. Arch Pharm Res. 2015;38(6):1178-1187.
- Poornima P, Weng CF, Padma VV. Neferine, an alkaloid from lotus seed embryo, inhibits human lung cancer cell growth by MAPK activation and cell cycle arrest. Biofactors. 2014;40(1):121-131.
- 14. Liu Y, Ma SS, Ibrahim SA, Li EH, Yang H, Huang W. Identification and antioxidant properties of polyphenols in lotus seed epicarp at different ripening stages. Food chem. 2015;185:159-164.
- Liao CH, Guo SJ, Lin JY. Characterisation of the chemical composition and in vitro antiinflammation assessment of a novel lotus (Nelumbo nucifera Gaertn) plumule polysaccharide. Food Chem. 2011;125(3):930-935.
- Liao CH, Lin JY. Lotus (Nelumbo nucifera Gaertn) plumule polysaccharide protects the spleen and liver from spontaneous inflammation in non-obese diabetic mice by modulating pro-/antiinflammatory cytokine gene expression. Food chem. 2011;129(2):245-252.
- Memon AA, Naz L, Shabbir S, Khan Z. The Hepato-Renal Protective Effect of Nelumbo nucifera Gaertn Seeds against Carbon Tetra Chloride Toxicity in Rats. Int J Pure App. Biosci. 2019;7(3):15-24.
- Itoh A, Saitoh T, Tani K, Uchigaki M, Sugimoto Y, Yamada J, Nakajima H, Ohshiro H, Sun S, Tanahashi T. Bisbenzylisoquinoline alkaloids from Nelumbo nucifera. Chem Pharm Bull. 2011;59(8):947-951.
- 19. Moro CF, Yonekura M, Kouzuma Y, Agrawal GK, Rakwal R. Lotus–a source of food and medicine: current status and future perspectives in context of the seed proteomics. Int J Life Sci. 2013;7(1):1-5.
- Akindele AJ, Ezenwanebe KO, Anunobi CC, Adeyemi OO. Hepatoprotective and in vivo antioxidant effects of Byrsocarpus coccineus Schum. and Thonn. (Connaraceae). J Ethnopharmacol. 2010;129(1):46-52.

- 21. Raajeswari PA, Meenakshi C. Antioxidant and Hepatoprotective Activity of Lotus (Nelumbo Nucifera) Seed Extract. Food Sci. Nutr. Tech. 2017;2(1), 2574-2701.
- 22. Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. Am J Clin Pathol. 1957;28(1):56-63.
- Tietz NW, Burtis CA, Duncan P, Ervin K, Petitclerc CJ, Rinker AD, Shuey D, Zygowicz ER. A reference method for measurement of alkaline phosphatase activity in human serum. Clin chem. 1983;29(5):751-761.
- 24. Sinha AK. Colorimetric assay of catalase. Anal Biochem. 1972;47(2):389-394.
- 25. Carlberg I, Mannervik B. Glutathione reductase. Methods Enzymol.-Vol. 113. United States: Academic press. 1985, pp. 484-490.
- Kono Y. Generation of superoxide radical during autoxidation of hydroxylamine and an assay for superoxide dismutase. Arch Biochem Biophys. 1978;186(1):189-195.
- 27. Okawa M, Kinjo J, Nohara T, ONO M. DPPH (1, 1-diphenyl-2-picrylhydrazyl) radical scavenging activity of flavonoids obtained from some medicinal plants. Biol Pharm Bull. 2001;24(10):1202-1205.
- Liu Y, Cao L, Du J, Jia R, Wang J, Xu P, Yin G. Protective effects of Lycium barbarum polysaccharides against carbon tetrachlorideinduced hepatotoxicity in precision-cut liver slices in vitro and in vivo in common carp (Cyprinus carpio L.). Comp Biochem Physiol Part C Toxicol Pharmacol. 2015;169:65-72.
- 29. Sheweita SA, Abd El-Gabar M, Bastawy M. Carbon tetrachloride-induced changes in the activity of phase II drug-metabolizing enzyme in the liver of male rats: role of antioxidants. Toxicology. 2001;165(2-3):217-224.
- Khan RA, Khan MR, Sahreen S. CCl4-induced hepatotoxicity: protective effect of rutin on p53, CYP2E1 and the antioxidative status in rat. BMC Complement Altern Med. 2012;12(1):1-6.
- 31. Morón ÚM, Castilla-Cortázar I. Protection against oxidative stress and "IGF-I deficiency conditions". Antioxidant enzyme. InTech Open. 2012:89.
- 32. Noureen F, Khan MR, Shah NA, Khan RA, Naz K, Sattar S. Pistacia chinensis: Strong antioxidant and potent testicular toxicity amelioration agent. Asian Pac j trop med. 2017;10(4):380-389.
- 33. Sohn DH, Kim YC, Oh SH, Park EJ, Li X, Lee BH. Hepatoprotective and free radical scavenging effects of Nelumbo nucifera. Phytomedicine. 2003;10(2-3):165-169.

219

34. Abdel-Kader MS, Abulhamd AT, Hamad AM, Alanazi AH, Ali R, Alqasoumi SI. Evaluation of the hepatoprotective effect of combination between hinokiflavone and Glycyrrhizin against CCl4 induced toxicity in rats. Saudi Pharm J. 2018;26(4):496-503.